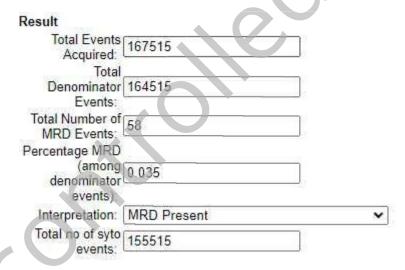
## Measurable Residual Disease for ALL by Flow Cytometry (MRD-ALL) Data Entry Redesign Guide

Participants can access an exercise by logging into their account on the participant hub and selecting the Measurable Residual Disease for ALL by Flow Cytometry programme. Participants can then select the current live exercise where they will see that some of the data entry fields have changed slightly.

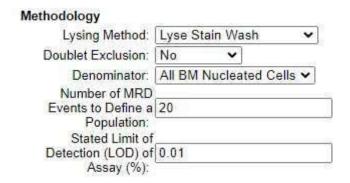
This section remains the same as before.

## Flow Cytometer: BD-FACSCanto II Flow MRD Group Affiliation: Non-Affiliated Panel set up (x-colour): 4 No. of tubes processed: 3 Nuclear dye correction used (syto): Yes

The results section now has two additional fields where participants are required to enter the Total Number of denominator events and a field where an interpretation of the results is required. The dropdown presents the option of MRD Absent, MRD Present and MRD Detected Below Quantifiable Limits.



A new methodology section has been introduced for participants to enter more information in relation to their assay technique. In this section you can enter information relating to lysing method, whether doublets are excluded form the analysis, the denominator used, the number of events used in your assay to define an MRD population and the stated Limit of Detection (LOD) of your assay.



There have been changes to the antigen section. Where we previously asked for antibody used, antibody manufacturer and fluorochrome, we now also require input of antibody clone, staining intensity and interpretation of the staining observed.

Antibody Manufacturer	Fluorochrome	Clone	Intensity	Result
Dako	FITC	123abc	Strong	Positive
BD Biosciences	PerCP	AD-1	Absent	Negative
Beckman Coulter	PerCP-CY5	104D2	Strong	Positive
Seralab	APC	*23d-1	Strong	Positive
In House	Chrome Orange	201	Weak	Positive
- 🗸 - Select a Manufactu 🛚	Select a Fluorochrc 🗸		- Select an Intens	ity V - Select - V
	Dako BD Biosciences Beckman Coulter Seralab In House	Dako FITC BD Biosciences PerCP Beckman Coulter PerCP-CY5 Seralab APC In House Chrome Orange	Dako         FITC         123abc           BD Biosciences         PerCP         AD-1           Beckman Coulter         PerCP-CY5         104D2           Seralab         APC         *23d-1	Dako         FITC         123abc         Strong           BD Biosciences         PerCP         AD-1         Absent           Beckman Coulter         PerCP-CY5         104D2         Strong           Seralab         APC         *23d-1         Strong           In House         Chrome Orange         2D1         Weak

Column 1 is **Antibody Used** – use the drop down to select this and if your antigen is not visible, please contact UK NEQAS LI to make the necessary amendments.

Column 2 Is Antibody Manufacturer - select the manufacturer from the dropdown list.

Column 3 is **Fluorochrome** – use the dropdown to select the fluorochrome used for the antibody.

Column 4 is **Clone-** Please enter the clone for the antibody used. This information is useful during data analysis, especially where there are discrepancies in the same antibody from different manufacturers. This is currently a free text box but for future exercises it is hoped that there will be a drop-down list as we build a collection of clones on our database.

Column 5 is Intensity- use the dropdown to select from a list of Absent, Weak and Strong.

Column 6 is **Result-** Use the dropdown to select whether your results are positive or negative for the selected antigen.

Once a panel has been saved down you will be able to select the same panel again for future exercises removing the need to enter all the information again. Simply select the panel from the previous exercise from the dropdown and hit 'Copy'. The panel may then be amended as required and saved.

	<del></del>	
Copy Antigens from previous panel:	SELECT	✓ Copy